

**1725-Pos****The E71A Mutation Alters Selective Ion Permeability in KcsA****Wayland W.L. Cheng**, Colin G. Nichols.

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The mechanism of selectivity in potassium channels has been extensively studied using the prokaryotic potassium channel, KcsA. Computational studies suggest that a glutamate-aspartate H-bond behind the selectivity filter in KcsA may play a role in determining the permeation properties of the channel. However, the mutant E71A, which disrupts this H-bond interaction and abolishes pH-dependent inactivation is reported to have either no effect on K<sup>+</sup> selectivity (1) or to increase K<sup>+</sup> selectivity (2) as measured by reversal potentials. Using an 86Rb<sup>+</sup> flux assay, WT KcsA exhibits strong K<sup>+</sup> selectivity, such that there are no measurable 86Rb<sup>+</sup> fluxes supported by Na<sup>+</sup> and Li<sup>+</sup>. In contrast, both Na<sup>+</sup> and Li<sup>+</sup> support significant 86Rb<sup>+</sup> fluxes in the E71A mutant, indicating an enhanced Na<sup>+</sup> and Li<sup>+</sup> permeability.

In eukaryotic inward rectifying potassium channels (Kir), the E71 equivalent residue is part of a glutamate-arginine salt bridge that, when disrupted dramatically reduces K<sup>+</sup> selectivity. KirBac1.1 is a prokaryotic channel that serves as a structural model of eukaryotic Kir, but contains an H-bond in the equivalent position, similar to KcsA. By patch-clamping giant liposomes, we show that KirBac1.1 is K<sup>+</sup>-selective (P<sub>Na</sub>/P<sub>K</sub> < 0.08) as measured by reversal potentials shifts, but, like KcsA E71A, shows significant Na<sup>+</sup> and Li<sup>+</sup>-driven 86Rb<sup>+</sup> fluxes. We also find that the KcsA E71A mutant, similar to WT KirBac1.1. This loss of stability in these channels may suggest that the differences observed in permeation result from a weakened interaction with ions at the selectivity filter. Studies to examine ion permeation in eukaryotic Kir channels by 86Rb<sup>+</sup> flux are ongoing.

1. H. Choi, L. Heginbotham, *Biophys.J.* 86, 2137 (2004).2. J. F. Cordero-Morales et al., *Nature Structural & Molecular Biology* 13, 311 (2006).**1726-Pos****Characteristic Frequency Analysis of Inward Rectifier Kir 2.1****John Rigby**, Steven Poelzing.

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INTRODUCTION: Impedance spectroscopy cannot distinguish between ion channel subtypes. We hypothesized that amplitudes of specific characteristic frequencies will correlate with the current amplitude passed by a specific ion channel subtype (characteristic frequency). We chose to test this hypothesis using the human inward rectifying potassium channel, Kir 2.1.

METHODS: IV-relationships were generated using a standard voltage step protocol (−140 to 0mV, 7mV steps) performed in whole-cell voltage clamp mode on HEK293 cells stably transfected with KCNJ2, which encodes Kir 2.1. Noise functions containing equal magnitudes of 1-15 kHz frequencies (amplitudes: 25, 50, 75 or 100mV) were inserted into each voltage step. The real component of the Fast Fourier transform (FFT) of the output signal was calculated with and without noise for each step potential. The magnitude of each frequency as a function of voltage step was correlated with the IV-relationship.

RESULTS: In the absence of noise (control), magnitudes of all frequencies correlated poorly ( $|R| < 0.15$ ) with the IV relationship. With noise, magnitudes of frequencies between 0.2-1 and 2-4 kHz demonstrated high negative ( $R < -0.9$ ) and positive correlation ( $R > 0.9$ ) respectively, with the IV-relationship. Two nodes of zero correlation were also found (1.39 ± 0.10 kHz and 8.49 ± 0.74 kHz). Increasing noise amplitude increased the absolute value of the correlation for the aforementioned frequencies without significantly changing the nodes of zero correlation.

CONCLUSIONS: These data suggest that the observed frequency response reflects current passing through Kir 2.1 channels. However it remains unknown whether any of these characteristic frequencies are unique to Kir2.1. Identifying characteristic frequencies of other ion channel subtypes could allow simultaneous measurement of multiple ionic currents.

**1727-Pos****Calcium Channels Exhibit Electric Field Dependent Valve-Like Behavior****James P. Barger**, Patrick F. Dillon.

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We propose a new model characterizing the valve nature of ion channels. There are four fundamental elements that any physiological valve must possess: a tube, a one-way gating mechanism, a gate-sensitive force, and a conducted substance. Macroscopic valves (heart or veins) have the gating mechanism attached to the tube. In our model of ion channels, the gating mechanism is the hydration state of the ion. A sufficient membrane electric field at the surface of an ion channel will strip water molecules from the ion, allowing the ion to

enter the channel. The electric field decays exponentially away from the membrane within several nanometers. If the ion channel extended too far from the membrane, negligible hydration stripping would occur, the hydration shell would remain around the ion, and the ion could not enter the channel. Our measurements of ion mobility in an electric field show that hydration stripping occurs at 400 V/cm, corresponding to 7 nm from the membrane. Calcium ion channels extend 4 nm externally from the membrane, and will have the ion hydration shells stripped from the ion at the channel entrance. Internally, the calcium ion channel extends 12 nm from the membrane. A stripped calcium ion will enter on the external side of the channel, but upon exiting on the internal side, will rehydrate and be unable to re-enter the channel, creating one-way flow. Thus, the calcium channels exhibit valve behavior, with the channel being the tube, the hydration shell the gating mechanism, the electric field the gate-sensitive force, and the stripped ion the conducted substance. This model can be extended to other ion channels. The macroscopic valves are thyroretic (tube-attached gate), while the ion channel valves are thyrofluidic (conductor-attached gate).

**1728-Pos****Selecting Ions by Size in a Calcium Channel: the Ryanodine Receptor Case Study****Dirk Gillespie<sup>1</sup>**, Le Xu<sup>2</sup>, Gerhard Meissner<sup>2</sup>.<sup>1</sup>Rush University Medical Center, Chicago, IL, USA, <sup>2</sup>University of North Carolina, Chapel Hill, NC, USA.

Calcium channels not only distinguish between ions of different charge (e.g., Ca<sup>2+</sup> vs. Na<sup>+</sup>), but also between of the same charge but of different size (e.g., Na<sup>+</sup> vs. K<sup>+</sup>). Size selectivity in calcium channels is analyzed in the ryanodine receptor (RyR) using a recent permeation model of RyR. This model describes ion permeation as electrodiffusion and ions as charged, hard spheres. RyR is modeled as five conserved negatively charged amino acids whose terminal carboxyl groups are very flexible. The model correctly reproduces experiments where three different monovalent cations compete for the pore at many different concentrations. Size selectivity occurs both because smaller ions fit into the crowded selectivity filter better and because they can screen the protein's negative side chains more effectively.

**1729-Pos****Insights from a Toy Model of Calcium Channels on Sieving Experiments and Eisenman Sequences****Daniel M. Krauss<sup>1</sup>**, Dirk Gillespie<sup>2</sup>.<sup>1</sup>Grinnell College, Grinnell, IA, USA, <sup>2</sup>Rush University Medical Center, Chicago, IL, USA.

A simplified model of a calcium channel is used to re-evaluate interpretations of both sieving experiments and Eisenman selectivity sequences in calcium channels. In the model channel, the carboxyl groups of the calcium channel selectivity filter are approximated as a homogeneous liquid of half-charged oxygens (two for each carboxyl group) separated from the bath by a semipermeable membrane that allows permeating ions into the selectivity filter, but does not allow the oxygens to leave the filter. Sieving experiments are usually interpreted with the logic that the physical diameter of a channel is equivalent to the largest particle that will go through that channel. However, our model has no radial geometric constraints, but still produces results that show net flux of ions quickly dropping to zero as the ions increase in diameter. These results indicate that forces like crowding of ions in the pore act on large ions that keep them from entering the channel. These forces are related to the pore diameter, but the results of the experiments should not be interpreted as indicating the pore diameter directly. We also used this simplified model to attempt to discern why the 11 Eisenman selectivity sequences are the only ones that have been observed. By altering the dielectric constant of the selectivity filter (and thereby the penalties for shedding waters of hydration), as well as oxygen concentration within the channel, we observed the circumstances under which each Eisenman sequence appears. We also observed a small number of non-Eisenman sequences.

**1730-Pos****Molecular Simulation of Ompf Channel in Salts of Divalent Cations: Molecular Insight on Charge Inversion****Marcel Aguilera-Arzo<sup>1</sup>**, Carles Calero<sup>2</sup>, Jordi Faraudo<sup>2</sup>.<sup>1</sup>Universitat Jaume I, Castelló de la Plana, Spain, <sup>2</sup>Institut de Ciència de Materials de Barcelona-CSIC, Bellaterra, Spain.

Extensive recent experimental and theoretical work has shown that the interaction of biologically relevant divalent cations (such as Mg<sup>2+</sup>, Ca<sup>2+</sup>) has surprising properties. One of the most fascinating and unexpected effect is the so-called charge inversion or charge reversal phenomenon: cations accumulate at the interface in excess of its own bare charge, thus inverting the effective